

# Identification of S-genotypes in almond progenies by NEPHGE and PCR

K. Kamali\*, J.M. Alonso\*\*, R. Socias i Company\*\*, A. Ebadi\*\*\* and M.R. Fattahi\*\*\*

\*IROST, No, 71. Forsat Street, Enghelab Ave., Tehran (Iran)

\*\*Unidad de Fruticultura, Centro de Investigación y Tecnología Agroalimentaria de Aragón (CITA), Av. Montañana 930, 50059 Zaragoza, (Spain)

\*\*\*Dept. Hort., University of Tehran, Karaj (Iran)

**Abstract.** Almond (*P. amygdalus* Batsch) shows a gametophytic self-incompatibility system controlled by a multiallelic locus, known as the locus S. Self-compatibility has been related to Sf allele presence and this trait has become a priority in the main almond breeding programs and the search for new cultivars is focused on the evaluation of desirable traits in autogamous seedlings. Traditionally, self-compatibility has been assessed in almond by laborious and time consuming methods, such as determination of fruit set in bagged branches or microscopic observation of pollen tube growth after self-pollination in laboratory conditions. Recently, molecular methods have been developed to assess the S-genotype, such as the identification of stylar S-RNases by NEPHGE (non equilibrium pH gradient electro focusing) and the use of conserved and specific PCR primers for the amplification of fragments from the different S alleles. In this research we applied these molecular methods for the S-genotype assessment in two almond breeding progenies, from the crosses of the self-compatible elite selection 'G-2-25' ( $S_{11}S_i$ ) of the CITA, as female parent, with two self-incompatible cultivars, 'Desmayo Langueta' ( $S_1S_{25}$ ) and 'Marcona' ( $S_{11}S_{12}$ ), as male parents. Although no discrepancies were found between the two methods, PCR was more suitable than NEPHGE for S-genotype assessment. PCR is easier to optimize, cheaper, more precise and reliable. It is also possible to assess the genotype sooner than with NEPHGE, as flowers are not required for the determination, allowing an earlier elimination of the seedlings. In the 'G-2-25' ( $S_{11}S_i$ ) x 'Marcona' ( $S_{11}S_{12}$ ) progeny, the ratio of genotypes was 42%  $S_{11}S_{12}$  and 58%  $S_iS_{12}$ , approaching the Mendelian laws of transmission in spite of the slightly higher proportion of self-compatible seedlings. In the other family, 'G-2-25' ( $S_{11}S_i$ ) x 'Desmayo Langueta' ( $S_1S_{25}$ ), four S-genotypes are possible, but the ratios obtained were 21%  $S_1S_{11}$ , 53%  $S_1S_i$ , 0%  $S_{11}S_{25}$ , and 26%  $S_iS_{25}$ . These results showed that pollen carrying the  $S_{25}$  allele had only a 26% fertilization success as compared to 73 of pollen carrying the  $S_i$  allele. The absence of  $S_{11}S_{25}$  seedlings may imply the expression of a homozygous lethal trait in these zygotes and the distortion of the Mendelian ratios.

**Keywords.** Self-compatibility – PCR Method – NEPHGE Method.

## Identification de génotypes-S chez les descendances d'amandiers par les méthodes NEPHGE et PCR

**Résumé.** L'amandier (*P. amygdalus* Batsch) montre un système d'auto-incompatibilité gamétophytique contrôlé par un locus multiallélique, connu comme locus S. L'auto-compatibilité a été reliée à la présence de l'allèle Sf et cette caractéristique est devenue une priorité pour les principaux programmes d'amélioration de l'amandier, et ainsi la recherche de nouveaux cultivars se focalise sur l'évaluation des caractères désirables pour les plants autogames. Traditionnellement, l'auto-compatibilité a été évaluée chez les amandiers par des méthodes laborieuses et prenant beaucoup de temps, telles que la détermination de la nouaison pour les branches ensachées ou l'observation microscopique de la croissance des tubes de pollen après auto-pollinisation en conditions de laboratoire. Récemment, des méthodes moléculaires ont été développées pour évaluer le génotype-S, telles que l'identification de S-RNases stylaires par NEPHGE (non equilibrium pH gradient electro-focusing) et l'utilisation d'amorces PCR conservées et spécifiques pour l'amplification de fragments provenant de différents allèles-S. Pour cette recherche nous avons appliqué ces méthodes moléculaires pour l'évaluation du génotype-S chez les descendances améliorées de deux amandiers, à partir des croisements d'une sélection d'élite auto-compatible 'G-2-25' ( $S_{11}S_i$ ) du CITA, comme parent femelle, avec deux cultivars auto-incompatibles, 'Desmayo Langueta' ( $S_1S_{25}$ ) et 'Marcona' ( $S_{11}S_{12}$ ), comme parents mâles. Bien que l'on n'ait pas trouvé de divergences entre les deux méthodes, la PCR était plus adéquate que NEPHGE pour l'évaluation du génotype-S. La PCR est plus facile à optimiser, moins onéreuse, plus précise et fiable. Il est également possible d'évaluer le génotype plus tôt qu'avec NEPHGE, étant donné qu'il n'est pas nécessaire d'obtenir des fleurs pour la détermination, permettant ainsi une

élimination précoce des plants. Dans la descendance de 'G-2-25' ( $S_{11}S_1$ ) 'Marcona' ( $S_{11}S_{12}$ ), le quotient des génotypes était de 42%  $S_{11}S_{12}$  et de 58%  $S_1S_{12}$ , se rapprochant des lois mendéliennes de transmission malgré la proportion légèrement plus élevée de plants auto-compatibles. Pour l'autre famille, 'G-2-25' ( $S_{11}S_1$ ) 'Desmayo Langueta' ( $S_1S_{25}$ ), quatre génotypes-S sont possibles, mais les quotients obtenus étaient de 21%  $S_1S_{11}$ , 53%  $S_1S_1$ , 0%  $S_{11}S_{25}$ , et 26%  $S_1S_{25}$ . Ces résultats montrent que le pollen portant l'allèle  $S_{25}$  avait une réussite de fertilisation d'uniquement 26% comparée à 73 pour le pollen portant l'allèle  $S_1$ . L'absence de plants  $S_{11}S_{25}$  pourrait faire penser à l'expression d'un caractère létal homozygote chez ces zygotes et à la distorsion des quotients mendéliens.

**Mots-clés.** Auto-compatibilité – Méthode PCR – Méthode NEPHGE.

---

## I – Introduction

Almond (*Prunus amygdalus batch* syn. *Prunus dulcis* [Mill] D.A. Webb) is one of the most important nut trees in the world. Main traditional almond cultivars are self-incompatible, However, there are some self compatible such as 'Touno', 'Supernova', 'Ginco', etc. (Raynaud and Grassely, 1985).

Self-incompatibility in almond is gametophytic and controlled by a single S-locus with multiple codominant alleles (Socias i Company *et al.*, 1976).

This trait is expressed in the style by special glycoproteins (S-RNases) that arrest the growth of pollen tubes in self-incompatible cultivars (Socias i Company *et al.*, 1976; Boskovic *et al.*, 1977).

Nowadays, using molecular methods is very important for identification of self-compatible and self-incompatible cultivars. In addition these methods are very reliable for determination of S-genotypes in almond. So far, it has been shown that there are about 27 self incompatible alleles ( $S_1, S_2, \dots, S_{27}$ ) and one self-compatible allele ( $S_?$ ).

Analysis of style ribonucleases (NEPHGE) and PCR methods are two important molecular methods to assess self compatibility in almond. The purpose of this research was the determination of S-genotypes in almond progenies, by using PCR and NEPHGE methods.

## II – Material and methods

### 1. Plant material

Thirty one almond progenies obtained from crossing of 'G-2-25' as a self-compatible cultivar with 'Desmayo Langueta' and 'Marcona', both as self-incompatible cultivars, were studied.

'G-2-25' is one of the most late blooming cultivars in Spain, as well as 'Desmayo Langueta' and 'Marcona' which are commercial early blooming spanish cultivars.

### 2. PCR method

We used several specific and conserved primers for identification of S-alleles in the almond progenies as follows:

(i)  $S_F$ - $S_R$  (Forward, GTGCCCTATCTAATTTGTTGAC) and (Reverse, GACATTTTTTTAGAAAGAGTG3) (Channuhtapipat *et al.*, 2003).

(ii) *conF*-*conR* (Forward,GTGCAACAATGGCCACCGAC) and (Reverse,TACCACTTCATGTAACAACACTGAG) (Channuhtapipat *et al.*, 2001, Tamura *et al.*, 2000).

(iii) *ASIII*-*AmyCSR*(Forward, TATTTTCAATTTGTGCAACAATGG) and Reverse, CAAAATACCACTTCATGTAACAAC) (Gómez and López, 2004).

### 3. PCR condition

Genomic DNA was extracted from 1 gr fresh leaves according to Gepts and Clegg method (1989). Long genomic DNA parts were amplified in a 25 µl reaction containing: Mmol (tris-HCl, pH=8), 100 Mm of each *dNTPs*, 1.5 mM MgCl<sub>2</sub>, 0.25 mM of each primers and 5 unit of *Taq DNA* polymerase. PCR program for *conF-conR* and *S<sub>i</sub>F-S<sub>i</sub>R* was an initial denaturation time of 3 min. at 94°C, followed by 34 cycles of 30 sec. denaturation at 95°C, 45sec annealing at 55°C and 1 min at 72°C with a final extension step of 10 min at 72°C. Cycling program for the *ASIII* and *AmyC5R* were: 5 min initial denaturation at 94°C, 30 cycles of 1 min denaturation at 94, 1 min of annealing at 53°C and 2 min of extension at 72°C, and then a 5 min final extension.

### 4. Gel electrophoresis

PCR product were run in 2% normal agaros with TAE Buffer at 100 V for 90 min, then gel stained by ethidoum bromid (1mg ml<sup>-1</sup>) and size of PCR products were determined with 1,2, 3 bp ladder.

NEPHGE (Non equilibrium pH gradient Electro focusing ): in this method, first of all, 120 styles were collected for each progenies at D stage (Boskovic *et al.*, 1997) , were put in 4 small tubes (1.5 ml Ependorf) 30 samples per each and stored at -80°C until next step (Boskovic *et al.*, 1997).

Extracted proteins from styles were separated electrophoretically on poly-acrylamide gel with 10 and 12% concentrations. It was run for 20 min at 100 V, 1 h at 150 V, 1 h at 100 V and 3 h at 400 V.

Later, gel were stained by toluiden blue for about 20 min. It was followed by immersing gel in distilled water for 12 h. Finally zymograms were compared (Boskovic *et al.*, 1997).

## III – Results and discussion

Results showed that both PCR and NEPHGE methods are suitable for identification of S-genotypes in almond. However, PCR method is easier. Regarding the use of several primers, we could identify all self-compatible genotypes from self-incompatible genotypes by using *S<sub>i</sub>F-S<sub>i</sub>R* primer, which produced 459 bp band for *S<sub>i</sub>* allele. *conF-conR* primer could identify *S<sub>ii</sub>* and *S<sub>i</sub>* alleles, whereas *ASIII* and *AmyC5R* could amplify *S<sub>i</sub>*, *S<sub>11</sub>* and *S<sub>12</sub>* alleles. However *conF-conR* and *ASIII* and *AmyC5R* could amplify *S<sub>i</sub>* alleles, but *S<sub>1</sub>* and *S<sub>25</sub>* alleles were only amplified with *ASIII* and *AmyC5R* primers. These results are in the same line as López *et al.*, 2004. Results obtained also showed that *conF-conR* could not amplify *S<sub>25</sub>* allele but it amplified *S<sub>i</sub>* allele.

The results obtained from NEPHGE method showed that this method needs more time compared to PCR. This method is more difficult and more expensive. However, its precision and accuracy are very high.

The ratio of S-genotypes obtained from crossing between 'Marcona' and 'G-2-25' cultivars, were according to Mendelian law. However, it was not exactly true for the crossing between 'Desmayo Larqueta' and 'G-2-25' cultivars, in which *S<sub>11</sub>S<sub>12</sub>* genotype was absent (21% *S<sub>1</sub>S<sub>11</sub>*, 53% *S<sub>1</sub>S<sub>i</sub>*, 0% *S<sub>11</sub>S<sub>25</sub>* and 26% *S<sub>25</sub>S<sub>i</sub>*).

These results showed that pollen carrying the *S<sub>25</sub>* allele had only a 26% fertilization success as compared to 73% of pollen carrying the *S<sub>i</sub>* allele.

In addition, the absence of  $S_{11}S_{25}$  seedlings may imply the expression of a homozygous lethal trait in these zygotes and the distortion of the Mendelian ratios.

## References

- Alonso J.M. and Socias i Company R., 2005a.** Identification of the  $S_3$  self-incompatibility allele in almond by specific primers. In: *Spain J. Agric. Res.*, 3, p. 296-303.
- Alonso J.M. and Socias i Company R., 2005b.** Differential pollen tube growth in inbred self compatible almond genotypes. In: *Euphytica*, 144, p. 207-213.
- Boscovik R., Tobutt K.R. Batlle I. and Duval H., 1997.** Correlation of ribonuclease zymograms and incompatibility genotypes in almond. In: *Euphytica*, 97, p. 167-176
- Boscovik R., Tobutt K.R., Duval H., Batlle I., Dicenta F. and Vargas F.J., 1999.** A stylar ribonuclease assay to detect self-compatible seedlings on almond progenies. In: *Theor. Appl. Genet.*, 99, p. 800-810.
- Boscovik R., Tobutt K.R., Batlle I., Duval H., Martínez-Gómez P. and Gradziel T.M., 2003.** Stylar ribonuclease in almond: Correlation with, and prediction of incompatibility genotypes. In: *Plant Breed.*, 122, p. 70-76.
- Channuntapipat C., Sedegly M. and Collins G., 2001.** Sequences of the cDNAs and genomic DNAs encoding the  $S_{11}, S_{7}, S_8$  and  $S_i$  alleles from almond *Prunus dulcis*. In: *Theor. Appl. Genet.*, 103, p. 1115-22.
- Channuntapipat C., Wirthensohn M., Ramessh S.A., Batlle I., Arus P., Sedegly M. and Collins G., 2003.** Identification of incompatibility genotypes in almond (*Prunus dulcis* Mill.) using specific primers based on the introns of the  $S$ -alleles. In: *Plant breed.*, 122, p. 164-168.
- López M., Mnejja M., Rovira M., Collins G., Vargas F.J., Arus P. and Batlle I., 2004.** Self-incompatibility genotypes in almond re-evaluated by PCR, stylar ribonucleases, sequencing analysis and controlled pollinations. In: *Theor. Appl. Genet.*, 109, p. 954-964.
- Socias i Company R., 1990.** Breeding self-compatible almond. In: *Plant Breed. Rev.*, 8, p. 313-338.
- Socias i Company R., 2001.** Different growth of almond pollen tubes in three environments. In: *Cahiers Options Méditerranéennes*, 56, p. 59-64.
- Socias i Company R and Alonso J.M., 2004.** Cross-incompatibility of 'Ferragnes' and 'Ferralise' and pollination efficiency for self-compatibility transmission in almond. In: *Euphytica*, 135, p. 333-338.
- Tamura M., Ushigima K., Sassa H., Hirano H., Tao R., Gradziel T.M. and Dandekar A.M., 2000.** Identification of self-incompatibility genotypes of almond by allele specific PCR analysis. In: *Theor. Appl. Genet.*, 101, p. 344-349.