

1 **First report of *Fusarium oxysporum* causing wilt and root rot in common borage**
2 **(*Borago officinalis*) in Spain**

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15 Common borage (*Borago officinalis* L.) is a horticultural species largely cultivated and
16 consumed in the center of Ebro valley in the Zaragoza province, northeast of Spain.
17 Over the last 10 to 15 years, greenhouse crops from different producers have been
18 affected by disease outbreaks characterized by symptoms such as severe yellowing and
19 wilting. Symptomatic plants' crown tissues and upper roots are covered with reddish-
20 brown to blackish-brown necrosis, which sometimes progresses up to the lower third of
21 the stem including leaf veins, and is followed by plant death. Severely infected plants
22 are stunted and eventually die, resulting in up to 70% crop loss. Small pieces (3-4 mm)
23 of symptomatic tissues from crown and roots were surface disinfected (0.5 min in 60%
24 ethanol and 2 min in 3% NaOCl), washed four times with sterilized distilled water,
25 plated onto potato dextrose agar (PDA) amended with streptomycin sulphate (0.5 g L⁻¹),
26 and incubated at 25°C in the dark for 1-3 days. *Fusarium*-like colonies were
27 consistently isolated and transferred to PDA and Spezieller Nährstoffarmer agar for
28 morphological characterization. Macroconidia were slender, pointed at the apexes,
29 fusoid-subulate or falcate, 3-5 septate in average, 48 (29-53) x 4.1 (3.5-4.5) µm. Aerial
30 microconidia were abundant, borne on short, lateral, narrowly lageniform

31 monophialides, ovoid, oval-ellipsoid or cylindrical, sometimes allantoid, mostly
32 aseptate 8 (5.2-17) x 3.1 (2.5-3.8) μm . Chlamydospores were globose, mostly
33 intercalary and not in chains. Isolates were tentatively identified as belonging to the
34 *Fusarium oxysporum* Schltdl. species complex (Booth, 1970). The internal transcribed
35 spacer region (ITS) and translation elongation factor-1 α (TEF-1 α) gene of isolate MYC-
36 1304 were PCR amplified using ITS1/ITS4 (White et al., 1990) and EF1-728F/EF1-
37 986R (Carbone and Kohn, 1999) primers, respectively, sequenced and deposited in
38 GenBank with accession numbers MK290853 (ITS) and MK332141 (TEF-1 α).
39 BLASTn analysis of the two sequences showed 99% and 100% homology with those of
40 *F. oxysporum* KU527806 (ITS) and LS479649 (TEF-1 α), respectively. Isolate MYC-
41 1304 was grown in 250 ml flasks containing potato sucrose medium for 3 days at 25°C
42 in the dark with constant agitation. Ten 20-day-old borage seedlings (cv ‘Movera’)
43 grown in trays with sterilized substrate, were removed, dipped into a suspension of 6 x
44 10⁶ conidia/ml for 2 min, and transferred to plastic pots with sterilized substrate. Three
45 non-inoculated plants dipped in sterile water were used as controls. Plants were
46 incubated in a growth chamber (25°C; 16/8 h photoperiod). Severe wilting and
47 yellowing accompanied by dry necrosis of the central veins of some leaves, followed by
48 plant death were observed 10 days post-inoculation. Non-inoculated controls remained
49 asymptomatic. The fungus was re-isolated and identified using ITS and TEF-1 α
50 sequences from all the inoculated plants. Both *F. oxysporum* and *F. solani* have been
51 cited as a cause of Viper’s Bugloss wilt in Iran (Nasr Esfahani and Monazzah, 2011;
52 Okhovvat et al. 2005), in spite that such common name refers to both *B. officinalis* and
53 *Echium vulgare* L. in Europe. Other *Fusarium* species (i.e. *F. avenaceum*) have also
54 been cited from common borage (Mulencko et al., 2008). To our knowledge, this is the
55 first report of *F. oxysporum* causing wilt and root rot of common borage in Europe. The

56 disease represents a serious threat to this local vegetable and its epidemiology. Seeds
57 and seedlings as well as their production substrate have been hypothesized as possible
58 carriers of infection propagules.

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