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8 **Intraspecific competition results in reduced evenness of *Tuber melanosporum* mating**
9 **type abundance from the nursery stage**

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27 **Abstract**

28 The highly-prized black truffle is a fungus mostly harvested in orchards planted with
29 mycorrhizal seedlings. It is an obligatory outcrossing fungus with a single *MAT* locus
30 containing two alternative mating-type idiomorphs. In the orchards, at the mycorrhizal level,
31 these mating types are frequently spatially segregated. Some studies found that this
32 segregation was pronounced from the nursery stage, whereas others did not find such a
33 marked segregation. Besides, information on the host tree species and nursery conditions used
34 in Spain, one of the main truffle-producing countries, are very scarce. In this study we
35 investigated the temporal dynamics of mating types in nursery seedlings of *Quercus ilex* and
36 *Quercus faginea*, as well as the influence of cultural conditions in the nursery. Our results
37 indicated that at the plant level, there was a trend for one of the mating types to dominate over
38 the other from the first to the second year in the nursery, in both host species and both nursery
39 conditions tested. However, this segregation process was not so sharp as previously reported.
40 Our results support the hypothesis that intraspecific competition results in reduced evenness
41 of mating type abundance from the nursery stage, although almost all seedlings maintained
42 both mating types and, at the seedling batch scale, the occurrence of both mating types was
43 roughly balanced.

44

45 **Keywords**

46 Truffle cultivation, *Tuber melanosporum*, mycorrhizal seedling, ectomycorrhiza, mating
47 types, MAT genes

48

49 **1. Introduction**

50 The European black truffle (*Tuber melanosporum*) is an ectomycorrhizal fungus highly
51 appreciated in *haute cuisine* due to its unique organoleptic qualities. Its wild production has
52 decreased in recent decades, so although black truffle grows in oak forests of southern
53 Europe, nowadays most of its production is harvested in orchards planted with seedlings
54 previously inoculated in nurseries under controlled conditions [1, 2]. Part of these plantations
55 have failed to produce truffles, due to factors such as unsuitable soils, dubious climate
56 conditions, poor quality of the mycorrhizal seedlings or inadequate cultural practices [3, 4].
57 Truffle cultivation is not completely domesticated yet, as the uncertainties around its life cycle
58 remain [5, 6]. Before and after truffle genome was sequenced, many studies have drawn
59 attention to the role of truffle sexuality in the productivity of truffle plantations [7–10].
60 *Tuber melanosporum* is a heterothallic organism, an obligatory outcrossing species with a
61 single *MAT* locus containing two alternative mating-type idiomorphs (*MAT1-1* and *MAT1-2*)
62 responsible for the mating process [7, 10, 11]. Mycorrhizae and ascospores of *T.*
63 *melanosporum* are formed by haploid cells, but truffle fruiting (sexual reproduction) requires
64 the concurrent presence of both mating types [11, 12]. During this mating process, the
65 mycelium from ectomycorrhizae behaves as the maternal partner, whereas ephemeral free
66 mycelia present in the soil act as paternal material [8, 12, 13].
67 In the nursery, where mycorrhizal seedlings are mostly produced using a spore suspension as
68 the inoculum, this ensures the presence of both mating types in the roots, since each fruitbody
69 is likely to produce spores of both mating types at roughly equal ratios [12, 14]. However,
70 once seedlings are planted in the field, it is frequently found a spatial segregation of the
71 mycorrhizal mating types, with host plants in adult *truffières* tending to sustain only one
72 truffle genet or multiple genets of the same mating type [8, 13, 15]. This situation does not

73 seem to happen during the first year in the nursery, just after seedlings are inoculated. Rubini
74 et al. [12] found that during the first year the abundance of mating types in the seedlings was
75 balanced, whereas in the second year they only found both mating types in half the seedlings
76 due to genet competition and displacement from the roots. However, other studies have not
77 found such a marked trend, with all the analyzed seedlings presenting both mating types [16,
78 17]. Information about this displacement process and about the possible influence of host
79 species and nursery cultural practices could be useful for improving productivity and
80 sustainability in truffle cultivation.

81 Although there are many species susceptible to mycorrhization by *T. melanosporum*, the
82 consolidation of the symbiosis in the field and the harvesting of truffles over the years occurs
83 mainly with oak species [1, 18, 19]. Therefore, in Spanish nurseries, *Quercus ilex* and
84 *Quercus faginea* are the most commonly used species, the latter being a faster-growing
85 species that produces more fine roots in the nursery [20]. The carbon that the fungus obtains
86 from the plant through these fine roots is a major trophic resource for which ectomycorrhizal
87 fungi compete [21]. Not only the host species, but also the environmental conditions and the
88 cultural practices in the nursery influence the number of root tips produced by the seedling
89 and the timing of fine-root production [22]. This could also influence the intraspecific
90 competition in truffle, since in the first year in nursery primary infection (from spore-
91 generated mycelium) seems to be the dominant process, while in the second year secondary
92 infection (from associated-to-mycorrhizae mycelium) gains weight, as suggested by the fact
93 that mycorrhizae from a given root branch are frequently genetically identical [23]. Finally,
94 the genetic diversity of the truffle fruitbodies used as inoculum and its dose could also play a
95 role, since they may influence the genetic diversity of the mycorrhizal population established

96 on the seedling roots, as suggested by genotyping studies in orchards inoculated with
97 mycelium and orchards outside the natural distribution area [16, 24].
98 In the present study, we investigated the temporal dynamics of *T. melanosporum* mating types
99 in nursery seedlings and the influence of host plant species and cultural practices on these
100 dynamics. We evaluated the relative abundance of mating types during the first and second
101 year in the nursery for the two most frequent host plants in Spanish truffle plantations: *Q. ilex*
102 and *Q. faginea*. We raised the seedlings in two different locations, using different substrates,
103 containers and inoculation doses (all of them within the standard cultural practices in
104 commercial nurseries), in order to obtain contrasting seedling characteristics that would
105 improve the generalizability of the results. We hypothesized that mating types would show a
106 more unbalanced distribution in older seedlings. We also hypothesized that *Q. faginea* and
107 seedlings with faster root growth would show a more balanced representation of mating types,
108 since a major trophic resource for which the mycorrhizal fungi individuals compete (the root
109 tips from which they receive the plant carbon) would be less limiting [25].

110

111 **2. Materials and methods**

112 *2.1. Experimental design*

113 A full factorial design was used to evaluate the effect of three independent variables (plant
114 species, time from inoculation and nursery) and their interactions on the relative abundance of
115 the mating types in the roots of the nursery seedlings. Two plant species (*Q. ilex* and *Q.*
116 *faginea*) were evaluated at two different moments (6 and 18 months from inoculation). The
117 seedlings were raised at two distant nurseries (El Toro and Graus) with different growth
118 medium, container volume and inoculum application rate. We did so in order to obtain
119 contrasting seedling characteristics that improve the generalizability of the results. A total of

120 96 seedlings (12 replicates for each of the 8 treatments) were produced, although only 73 of
121 them were finally included in the study due to seedling mortality or to extreme defects in
122 seedlings morphology (e. g. stem die-back or lack of fine roots): 8 for *Q. faginea* in Graus at
123 month 6, 10 for *Q. ilex* in El Toro at month 6, 10 for *Q. faginea* in El Toro at month 18, and 9
124 for all the remaining combinations of host tree, nursery and time from inoculation (Table S1).
125 The *T. melanosporum* fruitbodies used as inoculum (2 kg, more than 60 fruitbodies) were
126 harvested fresh and mature from plantations in Huesca province (northern Spain). They were
127 surface cleaned with a brush under cool water, surface sterilized by immersion in ethanol
128 (96%) and flamed, taxonomically identified by morphological features, sliced thin, air dried
129 under room conditions and homogenized with a coffee grinder. The oak acorns were acquired
130 from the Spanish provenance region *Sistema Ibérico*, surface sterilized with a 10% sodium
131 hypochlorite solution for 30 min, and germinated in January 2016 in a vermiculite tray. In
132 May 2016, when most seedlings had 6-8 leaves and had formed lateral roots, they were
133 removed from the tray, mechanically root-pruned at the tap root to eliminate defects when
134 they existed, inoculated and transplanted to containers (650 mL Quick-pot[®] in El Toro
135 nursery and 450 mL Full-pot[®] in Graus). Seedlings with malformations, poor development or
136 scarce fine roots were excluded. The inoculation was performed by root powdering with a
137 talcum powder (hydrated magnesium silicate) carrier, following Garcia-Barreda et al. [20].
138 The inoculum quantity was adjusted to obtain a rate of 2.5 g fresh truffle per seedling in El
139 Toro nursery and a rate of 1.0 g in Graus. The potting substrate in the El Toro nursery
140 consisted of 12:6:1 (v/v) calcareous sandy loam soil, base-fertilized *Sphagnum* white peat
141 (Kekkila[®] White 420W), and limestone coarse sand, while in Graus it consisted of 3:2:1:1
142 (v/v) calcareous clay loam soil, Prohumin[®] 5050 Substrate (50% *Sphagnum* black peat, 50%
143 *Sphagnum* white peat), limestone coarse sand and perlite.

144 The seedlings were maintained in the nursery greenhouse and sprinkle irrigated to saturation
145 2-3 times per week during summer and once a week during winter. Both nurseries are located
146 in wild truffle producing regions with Continental Mediterranean climate and a mean annual
147 temperature of 12.0-12.5°C, although the El Toro nursery is located at 1000 m a.s.l. in
148 Castellón province (eastern Spain) and the Graus nursery is located at 600 m a.s.l. in Huesca
149 province (northern Spain).

150

151 2.2. Data collection

152 For each plant species × nursery combination, half of the seedlings were randomly selected
153 and analyzed in October-November 2016 (6 months from inoculation) and the other half in
154 October-November 2017 (18 months). In each sampling, the stem height and the root-collar
155 diameter of seedlings were measured and the number of root tips per seedling, *T.*
156 *melanosporum* mycorrhizae per seedling and proportion of root tips colonized by *T.*
157 *melanosporum* were evaluated.

158 The mycorrhizal status was assessed through random sampling of roots. Fine roots (diameter
159 < 2 mm) were cut under water in portions with length < 1 cm and spread over a grid with 2 ×
160 2 cm square size. A 10% of the grid squares were randomly selected, and the root tips were
161 counted. With the aid of binocular and optical microscopes, tips were classified as non-
162 mycorrhizal or mycorrhizal, and the latter were classified as *T. melanosporum* or contaminant
163 morphotypes [26].

164 Sixteen *T. melanosporum* mycorrhizae were randomly selected in each plant, which is a
165 sample size similar to that of previous nursery studies for this bivalent sexual locus [12, 17].
166 With this purpose, 16 grid squares were randomly selected and one mycorrhiza was taken
167 from each of these squares. These tips were cleaned under the stereomicroscope using fine

168 forceps, placed in 0.2-mL sterile tubes containing 10 μ L of Extraction Solution (Sigma-
169 Aldrich, USA) and stored at -20 °C for further molecular analysis.

170 For mating type characterizations, genomic DNA was extracted from each individual
171 ectomycorrhiza: frozen tips were incubated for 10 min at 24°C and 10 min at 95 °C, following
172 Extract-N-Amp™ (Sigma-Aldrich, USA) recommendations. Then, 10 μ L of Dilution
173 Solution (Sigma-Aldrich, USA) were added and tubes were centrifugated at 6.000 rpm for 2
174 min. A PCR was arranged with primers P19 and P20 to amplify *MAT1-1* and primers P1 and
175 P2 for *MAT1-2* designed for *T. melanosporum* [11]. Each 12,5- μ l PCR reaction consisted of
176 2,5 μ L of 5x MyTaq™ Reaction Buffer Red (Meridian Biosciences, USA), 0,5 μ L bovine
177 serum albumin (Sigma-Aldrich, USA) at 10 mg mL⁻¹ [27], 0.25 μ L of each primer at 10 μ M,
178 7 μ L of ultra pure water to reach the volume mix of 18 μ L and 0,25 μ L of MyTaq™ DNA
179 Polymerase (Meridian Biosciences, USA). 1,25 μ L of genomic DNA were added for each
180 sample. The PCR reactions were performed in a Applied Biosystems (USA) 2720 thermal
181 cycler with the following thermal profile (adapted from Marozzi et al. [17]): initial
182 denaturation step of 5 min at 94 °C, followed by 40 cycles of denaturation at 94 °C for 30 s,
183 annealing at 60 °C for 30 s and extension at 72 °C for 45 s; and a final extension step at 72 °C
184 for 7 min. A *T. melanosporum* fruitbody was collected and genomic DNA was extracted from
185 its spores [28], to be used as a positive control in PCR to highlight the proper amplification of
186 both mating type genes in every reaction. All PCR experiments included a negative control
187 (no DNA template). Ten μ L of PCR products were run on 1.8% (w/v) agarose gel
188 electrophoresis stained with SYBR® Safe DNA Stain (Invitrogen, USA).

189 ITS regions of competing ectomycorrhizae (not *T. melanosporum*) DNA were sequenced
190 following the conditions detailed in Gómez-Molina et al. [29]. The quality of the obtained
191 sequences was assessed, and low-quality edges were removed with 4Peaks v1.7.2

192 (<https://nucleobytes.com/4peaks>). Fungal identification was carried out by searching highly
193 similar sequences in the GenBank and UNITE (<https://unite.ut.ee>) databases using the
194 megablast procedure and default settings [30].

195 2.3. Data analysis

196 The seedling root-collar diameter, stem height, number of root tips, number of *T.*
197 *melanosporum* mycorrhizae and percent root colonization by *T. melanosporum* were analyzed
198 with general linear models. When model assumptions (homogeneity of variance, normality
199 and linearity) were not met, the response variable was transformed. The proportion of
200 seedling roots colonized by each mating type (*MATI-1* vs. *MATI-2*) was analyzed with a
201 generalized (binomial) linear model. We also analyzed the proportion of root tips colonized
202 by the dominant mating type in each seedling (regardless of whether the dominant in that
203 seedling was *MATI-1* or *MATI-2*), in order to evaluate the evenness of mating type
204 representation (i. e. to measure how balanced is the abundance of the dominant and the non-
205 dominant mating type in a seedling). This proportion, which ranges from 0.5 (complete
206 evenness) to 1 (no evenness), was also analyzed with a generalized (binomial) linear model.
207 In the generalized linear models, the fit of the binomial error structure was assessed through
208 overdispersion, and when this assumption was not met, a quasibinomial error structure was
209 used. Least-squares means tests were used for post-hoc comparisons, with a $P = 0.05$
210 threshold for statistical significance.

211 A post-hoc correlation analysis was conducted to explore seedling characteristics that
212 influence the proportion of mycorrhizae colonized by the dominant mating type. With this
213 purpose, we calculated partial Pearson correlations between the proportion of the dominant
214 mating type and seedling characteristics, controlling for the age of the seedlings.

215 All analyses were conducted with R and the emmeans and correlation packages [31–33].

216

217 **3. Results**

218 The root-collar diameter of the seedlings was significantly affected by the interaction between
219 nursery and seedling age ($t = 3.50$, $P < 0.001$, Table S2, Fig. S1), with all seedlings

220 significantly increasing their size over time, although faster in the El Toro nursery (Table 1).

221 The stem height of the seedlings was significantly affected by the nursery ($t = 13.3$, $P <$
222 0.001), seedling age ($t = 4.4$, $P < 0.001$) and host species ($t = 2.1$, $P = 0.039$, Table S3, Fig.

223 S2), with height being higher in El Toro nursery, older seedlings and *Q. ilex* (Table 2). The

224 number of root tips was significantly affected by the interaction between nursery, age and

225 species ($t = 2.54$, $P = 0.014$, Table S4, Fig. S3), whereas the number of *T. melanosporum*

226 mycorrhizae was significantly affected by the nursery ($t = 5.83$, $P < 0.001$), seedling age ($t =$

227 7.21 , $P < 0.001$) and seedling species ($t = 2.43$, $P = 0.018$, Table S5, Fig. S4). Both root tips

228 and *T. melanosporum* mycorrhizae tended to be more abundant in older seedlings, in *Q.*

229 *faginea* seedlings and in the El Toro nursery (Tables 2, 3). The percent root colonization by *T.*

230 *melanosporum* was significantly affected by the interaction between nursery and seedling age

231 ($t = 2.02$, $P = 0.047$, Table S6, Fig. S5). Both nurseries showed high levels of colonization at

232 month 18, but only El Toro nursery presented high levels also at month 6 (Table 1). Besides

233 *T. melanosporum*, two other ectomycorrhizal morphotypes were found in the seedlings:

234 *Sphaerospora brunnea* in El Toro nursery (in 6.8% of seedlings) and *Telephora ellisii* in

235 Graus nursery (1.4% of seedlings).

236 More than 1000 ectomycorrhizae were analyzed and the mating type was determined in 862

237 of them, retrieved from a total of 73 seedlings (466 mycorrhizae with *MATI-1* and 396 with

238 *MATI-2*; Table S1). The generalized linear model analyzing the proportion of *MATI-1*

239 mycorrhizae did not show any significant effect of seedling species, age or nursery on the

240 proportion between mating types ($t = 1.78$, $P = 0.080$ for the seedling species; $t = 1.39$, $P =$
241 0.169 for the seedling age; and $t = -1.96$, $P = 0.054$ for the nursery, Table S7). All seedling
242 batches showed a predicted proportion that was not significantly different from 0.5, which
243 represents a 1:1 ratio, or very close to this value (Fig. 1).

244 The evenness of mating types abundance, computed as the proportion of root tips colonized
245 by the dominant mating type on each seedling, was significantly affected by seedling age ($z =$
246 2.56 , $P = 0.010$), with no significant effect of seedling species ($z = 1.29$, $P = 0.199$) or nursery
247 ($z = 0.27$, $P = 0.788$) (Table S8). At month 6, seedlings showed a predicted proportion that
248 was significantly higher than 0.5, but at month 18, this proportion significantly increased by
249 8%, this corresponding to a predicted change from a 2:1 to a 3:1 ratio between the dominant
250 and the non-dominant mating type in each seedling (Fig. 2). Despite these ratios, there were
251 only two seedlings (2.7%) in which just one mating type was found (one at month 6 and the
252 other at month 18).

253 Since the proportion of seedling mycorrhizae colonized by the dominant mating type showed
254 a significant time trend, a post-hoc correlation analysis was conducted to explore its
255 relationship with the seedling morphological characteristics. When controlling for seedling
256 age, the proportion of mycorrhizae colonized by the dominant mating type did not show any
257 significant correlation with the analyzed seedling characteristics (Table 4, Fig. S6).

258

259 **4. Discussion**

260 Our results indicate that the abundance of *MATI-1* and *MATI-2* in nursery seedlings was
261 overall balanced from the 6 to the 18 months after inoculation. They also indicate that, at the
262 plant level, there was a trend for one of the mating types to dominate over the other from the
263 first to the second year in the nursery. These results were consistent across two host species

264 and the contrasting growing conditions in two distant nurseries, agreeing with previous
265 studies on *Q. pubescens* [12] and *Carya illinoensis* [17], which used lower sample sizes.
266 Although our results broadly agree with those of Rubini et al. [12], they show some
267 differences regarding the extent of the process of displacement of the non-dominant mating
268 type. For seedlings in their second year in the nursery, Rubini et al. [12] only found
269 mycorrhizae of both mating types in 41% of the seedlings (this roughly corresponding to a 7:1
270 ratio between the dominant and the non-dominant mating type in each plant). Finding only
271 one mating type in the root system of a plant is common in the field, with older plants [12, 15,
272 16]. However, we found the two mating types in 97% of the seedlings in their second year,
273 whereas Linde and Selmes [16] and Marozzi et al. [17] found both mating types in all their
274 nursery seedlings. Interestingly, Rubini et al. [12] inoculated each seedling with the spores of
275 only one fruitbody, whereas we, Marozzi et al. [17] and probably Linde and Selmes [16] used
276 as inoculum the spores obtained from several fruitbodies, which is a common practice in
277 commercial nurseries. The quicker process of displacement of mating types that Rubini et al.
278 [12] found might be related to the lower genetic diversity of the inoculum and thus of the
279 mycorrhizal population initially established on the plants, which is a frequent outcome with
280 other organisms such as plants [34].

281 Other factors could also be the cause of this difference with Rubini et al. [12], such as
282 seedling species, inoculum dose, cultural practices in the nursery or size of seedlings (in so far
283 as size may reflect the abundance of root tips and/or the timing of fine root production).

284 However, in our study we did not find differences between *Q. ilex* and *Q. faginea*, despite the
285 fact that the latter produces more root tips and thus more mycorrhizae are formed [20].

286 Regarding the inoculum dose and cultural practices, previous studies do not detail such
287 factors, thus precluding comparisons. However, in our study we did not find differences

288 between the two nurseries, despite the fact that in one of them plants formed more root tips
289 and mycorrhizae, especially in the first year. After controlling for the effect of plant age, no
290 correlation was found between the abundance of root tips and the evenness among mating
291 types. Our results suggest that, for the range of root tip abundance in our study, this variable
292 does not condition the competition and displacement process between mating types, in spite
293 of root tips being a major trophic resource for mycorrhizal fungi [25].

294 The absence of a relationship between the abundance of the trophic resource (root tips) and
295 the intensity of the competition process (with the mating type evenness as a proxy) raises the
296 question of which is the trigger of the displacement process. Rubini et al. [12] results suggest
297 that differences in mating type evenness do not come from differences in the proportions of
298 mating types in the spore inoculum or in the ability of mating types for primary infection,
299 since they inoculated each seedling with spores of only one fruitbody and found a roughly
300 balanced occurrence of mating types in seedlings during their first year in the nursery. This
301 points to some process related with secondary infection (from other mycorrhizae and
302 associated mycelium), which gains importance in the second year in the nursery. Rubini et al.
303 [12] and Selosse et al. [5] hypothesized that the competition for root tip colonization is related
304 to a self/nonself recognition mechanism linked to the *MAT* locus. In vitro dual cultures of
305 genetically different *T. melanosporum* strains showed that hyphae from different strains did
306 not form anastomoses while hyphae from the same strain frequently did, suggesting a
307 vegetative incompatibility based on a mechanism acting before hyphal fusion [35]. In any
308 case, it is likely that this process of self/nonself recognition is not dependent on root tip
309 abundance.

310 In truffle orchards, the process of mating displacement from the plant roots seems to intensify.
311 In young plantations, Linde and Selmes [16] observed that only half of the plants maintained

312 mycorrhizae of the two mating types, whereas in adult orchards and wild *truffières* Murat et
313 al. [15] and Rubini et al. [12] found that most host trees had only one mating type in their
314 roots. It would be interesting to study whether the number of mycorrhizae formed in the
315 nursery, their genetic diversity, or the mating type evenness in the nursery can influence the
316 process of spatial segregation of mating types observed once the seedlings are planted in the
317 field. In any case, this spatial segregation at the mycorrhizal level does not seem to be so
318 marked at the level of free mycelia in the soil (some of which act as parental material in
319 mating), as suggested by recent research assessing both types of mycelia together [9, 36].
320 To conclude, at the seedling batch level we found a roughly balanced representation of the
321 two mating types in the mycorrhizae of seedlings inoculated with *T. melanosporum* until
322 month 18 after inoculation. Despite this, at the plant level we found a trend for one of the
323 mating types to dominate over the other during the second year in the nursery. Together, these
324 findings support the hypothesis that intraspecific competition results in reduced evenness of
325 mating type abundance from the nursery stage, corroborating that this process begins in the
326 second year, when the importance of secondary infection increases with respect to primary
327 infection. We did not find differences in this process related to host species (*Q. ilex* versus *Q.*
328 *faginea*), to cultivation conditions in the nursery or to seedling size. Our study corroborates,
329 with a larger sample size, the results of previous studies, extending them to the more frequent
330 host plants for *T. melanosporum* in Spain and connecting these results with cultivation
331 conditions and seedling characteristics, thus providing helpful information for commercial
332 nurseries producing mycorrhizal seedlings.

333

334 **Data Availability**

335 The datasets generated during the current study are available from the corresponding author
336 on reasonable request.

337

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451 **Statements and Declarations**

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456 **Author Contributions**

457 E.G.-M., S.S. and S.G.-B. conceived and designed the study. Funding was secured by E.G.-
458 M. and S.S. Laboratory protocols were conducted by E.G.-M., M.P.-P. and S.S. The data were
459 analyzed by S.G.-B. The manuscript was initially written by E.G.-M., along with significant
460 contributions from S.S. and S.G.-B.

461 **Competing Interests**

462 The authors declare no competing interests.

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465 **Table 1.** Effect of the interaction between nursery and seedling age on root-collar diameter of
 466 seedlings and percent root colonization by *T. melanosporum* (mean predicted values and 95%
 467 confidence intervals). Letters within each column indicate significant differences (least-
 468 squares means, $\alpha = 0.05$, $n = 73$).

	Root-collar diameter (mm)	Percent root colonization ¹
El Toro		
6 months	4.0 (3.6, 4.3)c	35.4 (28.0, 43.7)a
18 months	6.5 (6.1, 6.8)a	37.3 (29.7, 45.8)a
Graus		
6 months	3.4 (3.0, 3.7)c	11.8 (7.5, 17.1)b
18 months	4.7 (4.4, 5.0)b	29.1 (22.2, 36.9)a

469 ¹ Variable square-root transformed

470

471 **Table 2.** Effect of the host plant, the nursery and seedling age (main effects) on the height of
 472 the plant stem and the number of *T. melanosporum* ectomycorrhizae (ECM) per seedling
 473 (mean predicted values and 95% confidence intervals). For each predictor variable (host
 474 species, nursery, seedling age), letters indicate significant differences (least-squares means, α
 475 = 0.05, $n = 73$).

	Stem height (cm) ¹	No of truffle ECM ($\times 10^3$) ²
Host species		
<i>Q. ilex</i>	17.6 (16.0, 19.4)a	1.4 (1.1, 1.6)b
<i>Q. faginea</i>	15.3 (13.9, 16.8)b	2.2 (1.9, 2.5)a
Nursery		
El Toro	25.8 (23.5, 28.3)i	2.7 (2.3, 3.0)i
Graus	10.4 (9.5, 11.5)j	1.0 (0.8, 1.2)j
Seedling age		
6 months	14.1 (12.8, 15.6)q	0.6 (0.5, 0.8)q
18 months	19.0 (17.3, 20.9)p	3.4 (3.0, 3.8)p

476 ¹ Variable log-transformed

477 ² Variable square-root transformed

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481 **Table 3.** Effect of the interaction between nursery, host plant and seedling age on the number
 482 of root tips per seedling (mean predicted values and 95% confidence interval). Different
 483 letters indicate significant differences (least-squares means, $\alpha = 0.05$, $n = 73$).

	No of root tips ($\times 10^3$) ¹
<i>Q. ilex</i> El Toro	
6 months	2.42 (1.78, 3.16)d
18 months	9.59 (8.20, 11.10)b
<i>Q. ilex</i> Graus	
6 months	1.40 (0.90, 2.00)d
18 months	5.46 (4.42, 6.61)c
<i>Q. faginea</i> El Toro	
6 months	5.47 (4.43, 6.62)c
18 months	15.5 (13.8, 17.3)a
<i>Q. faginea</i> Graus	
6 months	2.01 (1.36, 2.77)d
18 months	12.0 (10.4, 13.7)b

484 ¹ Variable square-root transformed

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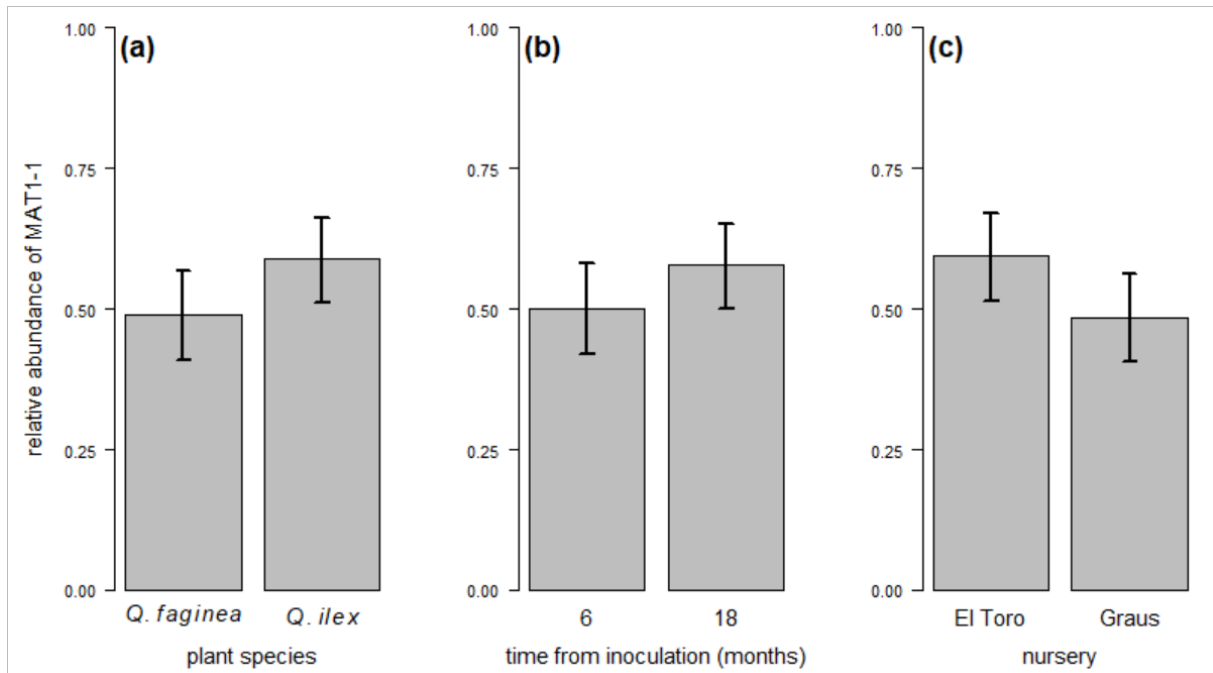
486 **Table 4.** Partial Pearson correlations between the characteristics of nursery seedlings and the
 487 evenness of mating types abundance (computed as the proportion of seedling mycorrhizae
 488 colonized by the dominant mating type in the seedling, square-root transformed), after
 489 controlling for seedling age.

	No of seedlings	Partial correlation	P-value
Root-collar diameter	73	-0.03	0.80
Stem height ¹	73	-0.14	0.24
Number of root tips ²	73	0.09	0.47
Number of <i>T. melanosporum</i> mycorrhizae [‡]	73	0.10	0.40
Percent roots colonized by <i>T. melanosporum</i> [‡]	73	-0.003	0.98

490 ¹ Variable log-transformed

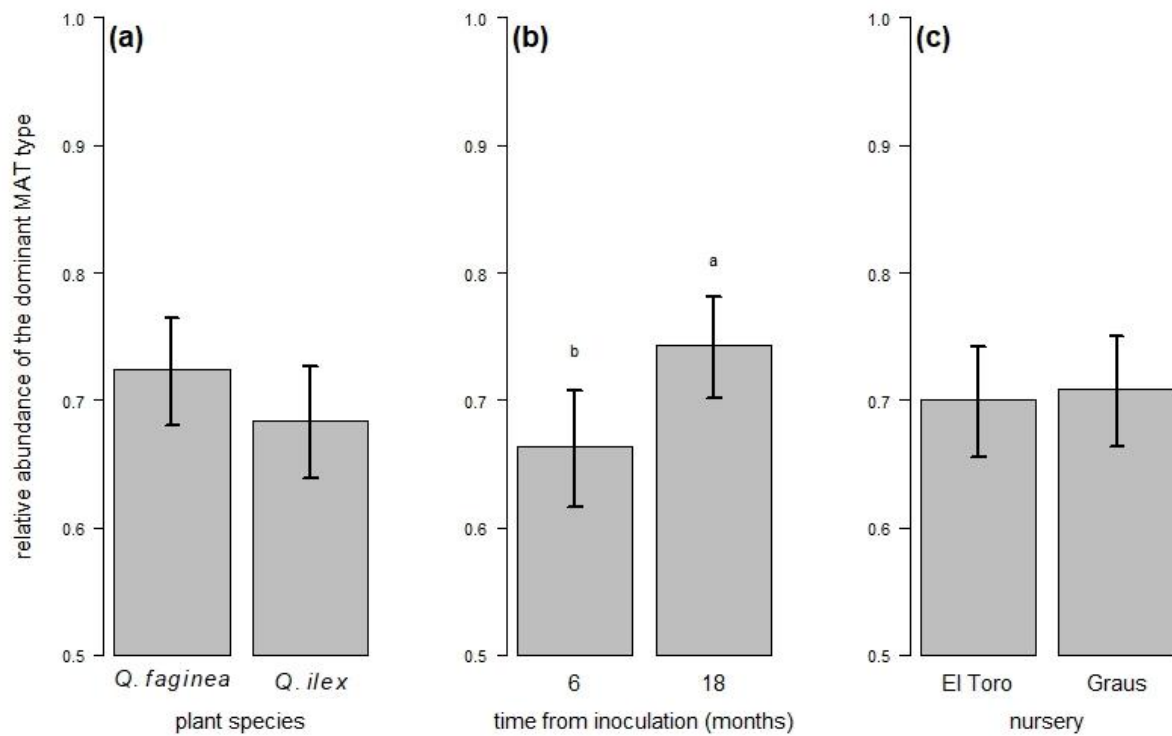
491 ² Variables square-root transformed

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Figure 1. Relative abundance of *MAT1-1* in the ectomycorrhizae of the nursery seedlings (mean predicted values and 95% confidence intervals, $n = 73$), according to seedling species (a), time from inoculation (b) and seedling batch (c). Overlapping of the confidence intervals indicates lack of significant differences according to the least-squares means procedure.



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Figure 2. Proportion of seedling mycorrhizae colonized by the dominant mating type in the seedling (mean predicted values and 95% confidence intervals, $n = 73$). A value of 1 means no evenness and a value of 0.5 means complete evenness between the two mating types. (a) Differences between seedling species. (b) Relationship with time from inoculation. (c) Differences between the two nursery batches produced. Letters indicate significant differences according to the least-squares means procedure ($\alpha = 0.05$).