- NOTICE: this is the authors' version of a work that has been accepted for publication in **Plant**
- 2 and Soil. Changes resulting from the publishing process, such as editing, corrections,
- 3 structural formatting, and other quality control mechanisms may not be reflected in this
- 4 document. Changes may have been made to this work since it was submitted for publication.
- 5 The final publication is available in Plant and Soil:
- 6 https://doi.org/10.1007/s11104-023-06106-3

7

- 8 Intercropping of aromatic plants in truffle orchards: short-term effect on extraradical
- 9 truffle mycelium and aromatic plant growth

10

- 11 Sergi Garcia-Barreda <sup>a,b\*</sup>, Juliana Navarro-Rocha <sup>a,b</sup>, Eva Gómez-Molina <sup>c</sup>, Vasiliki Barou <sup>d</sup>,
- 12 María Ángeles Sanz <sup>e,b</sup>, Sergio Sánchez <sup>a,b</sup>, Javier Parladé <sup>d</sup>

13

- <sup>a</sup> Departamento de Ciencia Vegetal, Centro de Investigación y Tecnología Agroalimentaria de
- 15 Aragón (CITA). Avenida de Montañana 930, Zaragoza 50059, Spain.
- b Instituto Agroalimentario de Aragón-IA2 (CITA-Universidad de Zaragoza), 50018
- 17 Zaragoza, Spain
- <sup>c</sup> Centro de Investigación y Experimentación en Truficultura (CIET), Diputación Provincial
- 19 de Huesca. Polígono Fabardo s/n, 22430, Graus, Spain.
- 20 d Protecció Vegetal Sostenible, Institut de Recerca i Tecnologia Agroalimentàries (IRTA).
- 21 Ctra. Cabrils km 2, E-08348 Cabrils, Spain.
- <sup>e</sup> Área de Laboratorios de Análisis y Asistencia Tecnológica, Centro de Investigación y
- 23 Tecnología Agroalimentaria de Aragón (CITA). Avda. Montañana 930, 50059 Zaragoza,
- 24 Spain.
- \* Corresponding author: sgarciaba@cita-aragon.es

26

- 27 ORCID
- 28 Sergi Garcia-Barreda: 0000-0002-7248-234X, Juliana Navarro-Rocha: 0000-0001-7975-
- 29 9340, Eva Gómez-Molina: 0000-0002-2664-8484, Vasiliki Barou: 0000-0003-2939-2426,
- 30 María Ángeles Sanz: 0000-0002-4513-3371, Sergio Sánchez: 0000-0003-4331-9794, Javier
- 31 Parladé: 0000-0002-0867-3280

#### Abstract

33

57

Background and Aims 34 Intercropping of truffle-producing trees with aromatic plants is used to improve profitability 35 of truffle orchards during the initial 4-7 years. However, after that period the viability of this 36 system is challenged by the appearance of *brûlés*, an area around host tree characterised by 37 scarce plant cover where the fungus exhibits allelopathic activity. We aimed to investigate the 38 ecological interactions between these crops and between their associated mycorrhizal fungi in 39 adult truffle plantations. 40 Methods 41 We simulated two intercropping systems, truffle oak – lavender and truffle oak – rosemary in 42 43 their adult stage. We analysed and compared aromatic plants and soil samples inside and outside the *brûlés* during the first year of the aromatic plants in the field. 44 45 Results We found a strong negative relation of *brûlés* with the growth of the aromatic plants, although 46 not a decrease in the arbuscular mycorrhizal colonization of their roots. The essential oil yield 47 and composition of aromatic plants was affected by brûlés. The extraradical truffle mycelium 48 49 was not significantly affected by the presence of aromatic plants. 50 Conclusions The growth and yield of aromatic plants was impaired during their first year growing in 51 brûlés, whereas no negative effect of aromatic plants on truffle fruiting potential was found. 52 53 The study improves our understanding of the mechanisms influencing the viability of the truffle tree – aromatic plant intercropping and the possible technical challenges. 54 55 **Keywords**: Tuber melanosporum, Lavandula × intermedia, Salvia rosmarinus, intercropping, 56

extraradical mycelium, medicinal and aromatic plants

### Introduction

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

Agricultural lands in the Mediterranean basin face important challenges due to climate and societal changes, such as soil erosion, reduced water availability, and degradation of soil organic matter and the associated microbial communities (De Franchis 2003; Lagacherie et al. 2018). In this context, arboriculture has a high potential to offer ecosystem services such as enhancement of farmers' income and carbon sequestration, although in many cases there are conflicts between provisioning and regulating services, frequently linked to overfertilisation, rainwater runoff due to reduced groundcover or cultivation practices disrupting interactions responsible for pest control and pollination (Brunori et al. 2020; Demestihas et al. 2017, 2019). Intercropping of orchards with medicinal and aromatic plants (MAPs) could mitigate these conflicts through increasing agricultural production, carbon sequestration, soil water retention, soil biodiversity, populations of pollinators and control of pests and diseases (Chen et al. 2014; Durán-Zuazo et al. 2008; Morugán-Coronado et al. 2020; Song et al. 2010; Zhang et al. 2021). The potential benefits of intercropping with MAPs could be particularly interesting for truffle cultivation. The prized black truffle (Tuber melanosporum Vittad.) grows and fruits below ground in ectomycorrhizal (ECM) association with tree roots. The productivity and sustainability of truffle orchards depends on the ecological relationships of truffle with soil components, host trees and soil microbial communities (Benucci and Bonito 2016; Le Tacon et al. 2013; Mello and Balestrini 2018; Splivallo et al. 2011). These relationships indicate the potential for agroecological approaches that integrate the diverse organisms interacting in the oak – truffle agroecosystem (Aumeeruddy-Thomas et al. 2016). Intercropping with MAPs could be an environmentally beneficial alternative (Schneider-Maunoury et al. 2020; Taschen et al. 2020), but the first step would be to develop farming practices harmonizing the association between these crops. A few experiences of intercropping of lavender (Lavandula

83 × intermedia Emeric ex Loisel.) and truffles exist in southern Europe (Martin-Chave 2019), although the scientific information on them is still very scarce (Geoffroy et al. 2018). 84 In truffle – lavender intercropping plantations, lavender is commonly planted in rows between 85 truffle tree rows (Martin-Chave 2019). During the pre-productive stage of the truffle 86 plantation – the first 4–7 years from tree plantation – the root systems of the crops do not 87 overlap and intercropping seems easily feasible. However, from year 4–7 an area 88 characterised by scarce plant cover (the so-called brûlé) is developed around the truffle trees 89 (González-Armada et al. 2010; Streiblová et al. 2012). The biological mechanism behind the 90 brûle formation has not been completely determined, but it likely involves competition among 91 92 roots, allelopathic activity and/or root parasitism by T. melanosporum mycelium (Pacioni 93 1991; Plattner and Hall 1995; Schneider-Maunoury et al. 2018). To the extent that *brûlés* may affect MAP growth, they could jeopardise the productivity of the MAP crop or its aromatic 94 95 profile, which is sensitive to other stress factors such as drought (Chrysargyris et al. 2016; Kulak 2020). This could be particularly problematic when MAPs are recently planted. 96 Plantations of lavender and rosemary (Salvia rosmarinus (L.) Schleid.) have a lifespan of 8-97 10 years, after which MAP rows must be replanted if the intercropping is going to be 98 99 maintained (Fanlo et al. 2009). 100 On the other hand, most truffle fruitbodies are harvested within brûlés, with the extraradical 101 truffle mycelium being more abundant in productive than non-productive brûlés (Parladé et al. 2013; Queralt et al. 2017). As brûlés stretch and reach a MAP row, the spread of truffle 102 103 mycelium could be hindered by the competition with MAP roots and their associated fungi, which are arbuscular mycorrhizal (AM) fungi in the case of lavender and rosemary. Previous 104 105 studies have frequently found a negative relationship between ECM and AM fungi (Chen et al. 2000; Knoblochová et al. 2017). 106

In this study, we explore the effect of intercropping on the growth of MAPs and on the abundance of truffle mycelium in the soil. Since the main challenges for the compatibility of these crops are likely to happen once the *brûlés* appear, and especially when MAPs are replanted, we studied these effects during the first vegetative period after planting MAPs in a ten-year-old truffle orchard with formed brûlés. We evaluated the survival, growth and essential oil characteristics of two common MAP species (lavender and rosemary) within the brûlé, as well as their root colonization by AM fungi. We compared them to MAPs planted outside the *brûlé*, where contact between tree and MAP roots is much more limited or null, just like in pre-productive truffle plantations. We also evaluated the effect of MAP occurrence on the abundance of T. melanosporum extraradical mycelium. Regarding the effect of intercropping on MAPs, we hypothesised that in the *brûle* the higher abundance of T. melanosporum mycelium would be associated with reduced survival and growth of MAPs and lower colonization of MAP roots by AM fungi. We also hypothesised that plants growing within the *brûlé* would suffer higher stress levels that would be associated with higher yields of essential oils (the main product of the cultivation of these MAPs) and altered volatile compounds profiles, since this is frequently the case for other stress factors such as drought (Chrysargyris et al. 2016; Kulak 2020; Sarmoum et al. 2019). Regarding the effect of intercropping on truffle, we hypothesised that the abundance of *T. melanosporum* mycelium would be impaired by the presence of MAPs, thus limiting the long-term compatibility of truffle – MAPs intercropping. We assessed two MAP species (lavender and rosemary) to test whether the truffle – MAP compatibility is dependent on the MAP species used.

128

129

130

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

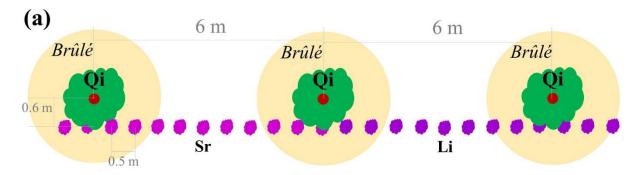
### **Materials and methods**

Experimental site and design

The experiment was conducted in the truffle orchard of Centro de Investigación y 131 Experimentación en Truficultura (CIET) in Graus (Huesca province, north-eastern Spain, 42° 132 12.1' N, 0° 22.4' E, 520 m. a.s.l.). The climate is Continental Mediterranean, with a mean 133 annual temperature of 12.2 °C and a mean annual rainfall of 680 mm. The soil is calcareous, 134 with pH 8.2, and silt-loam texture, developed on Tertiary lutite/sandstone. The site lies within 135 the natural distribution area of *T. melanosporum* (Garcia-Barreda et al. 2019). 136 A small, homogeneous surface of this plantation (ca. 400 m<sup>2</sup>) was used as experimental plot 137 in our study. This plot was planted in 2011 with *Quercus ilex* L. subsp. *ballota* (Desf.) Samp. 138 seedlings inoculated with T. melanosporum. The seedlings were produced in a commercial 139 140 nursery and the root colonization by T. melanosporum was evaluated following the INIA-141 Aragón method (Andrés-Alpuente et al. 2014), and checked to be at least 30% of the total short roots, with less than 1% of Sphaerosporella brunnea (Alb. & Schwein.) Svrcek & 142 Kubicka. The seedlings were planted at a density of 278 trees ha<sup>-1</sup> (6  $\times$  6 m). The soil has 143 been shallowly tilled once a year in early spring and afterwards the weeds have been 144 controlled (when necessary) by hoeing around the trees in spring and autumn. Since the sixth 145 year of the plantation, the trees have been biennially pruned. 146 147 In April 2021, the experimental plot was divided in two blocks. In the first block, 41 lavender 148 (clone super) were planted. In the second block, 81 rosemary seedlings were initially planted, although a small area was discarded due the presence and digging activity of *Microtus arvalis* 149 Pallas, thus leaving for the study 67 seedlings in the rosemary block. These two 150 151 Mediterranean, perennial shrubs are amongst the most common MAPs cultivated in the calcareous, dry regions of Spain (More et al. 2010). They are commercially cultivated to 152 153 obtain essential oil, fresh or dried sprigs. They are usually harvested by mechanically cutting flowering stems and thus crop yield is largely dependent on annual shoot growth (More et al. 154 2010). Following the common practice with these MAPs, the seedlings were planted in a row 155

with 50 cm distance between plants (Fig. 1). The row of MAPs was 60 cm apart from the row of truffle trees, so that 36% of the MAP seedlings were located within a truffle *brûlé* (at distances ranging 0.5-1.3 m from the truffle tree trunk) and 64% outside *brûlés* (at distances ranging 1.6-3.1 m). The former represents a situation in which crops could potentially interfere with each other, whereas the latter represents a situation in which the probability of this happening is much lower (Fig. 1). The *brûlés* of the experimental trees presented mean radius 1.0–1.5 m around the host tree trunk.

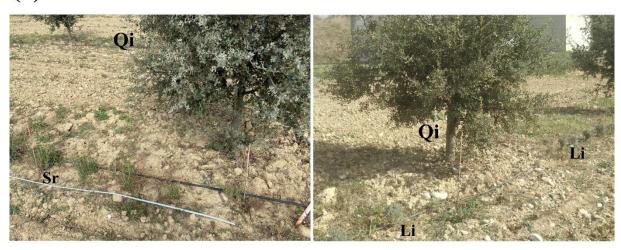
Following the common practice, lavender was planted bare root, whereas rosemary was grown potted in the CIET nursery and then planted with its root ball. The MAP seedlings



were drip watered once each 10-12 days (10 litres per seedling) during two months after

plantation, to encourage their survival and rooting.

(b)



**Figure 1**. General layout diagram of the experimental design (a) and overall appearance of the experimental plot six months after plantation. Qi: *Quercus ilex*, Sr: *Salvia rosmarinus*, Li: *Lavandula* × *intermedia*.

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

169

170

171

Plant measurement and mycorrhizal sample collection In October 2021, six months after planting, the survival and shoot height of all the MAPs (n = 108) were measured. Then, four truffle trees in the lavender block and four in the rosemary block were selected. For each tree, four MAPs were selected, two of them within the brûlé and two outside, to assess the effect of the *brûlé* on the MAPs (sample size: 32 MAPs). The plants were thoroughly uprooted and soil from the rhizosphere was collected by gently shaking the plant roots, for subsequent quantification of *T. melanosporum* extraradical mycelium. Then, the fine roots of each MAP were cut and kept in moist conditions at 4°C, for subsequent quantification of AM fungi colonization. The sample size for AM fungi colonization was reduced to 12 MAPs (three truffle trees for each MAP species and two MAPs per tree). Finally, the shoots of the 32 MAPs were dried to constant weight at 80 °C. In the eight truffle trees selected (four with lavender and four with rosemary) a bulk soil sample from the *brûlé* was taken. It consisted of a composed sample from three cylindrical cores (20 cm depth, 3.2 cm diameter), at 0.5-1 m distance from the truffle tree trunk. In each brûlé, two samples of soil from MAP rhizosphere were taken, as described above. Rhizosphere samples were compared to the bulk soil to assess the effect of MAP presence on the abundance of *T. melanosporum* extraradical mycelium (sample size: 24). For each tree, one bulk soil sample from outside the *brûlé* (at 2.5-3 m from the truffle tree trunk) was taken with the same methodology, as a control. Finally, as a complementary analysis, we sampled dead MAPs in the *brûlés* to compare the abundance of T. melanosporum extraradical mycelium in their rhizosphere with that of living

194 MAPs in the same brûlé. Four dead MAPs were sampled, each one in a different brûlé, together with two living MAPs in each brûlé, for a total sample size of 12. 195 196 197 Extraradical mycelium of black truffle The soil samples were air-dried at 30 °C and sieved through a 2-mm mesh. DNA extraction 198 was performed on the fine soil fraction using the Soil DNA Isolation Plus Kit (Norgen Biotek 199 Corp., Thorold, ON, Canada) following manufacturers' instructions. Specific quantification of 200 soil mycelium was carried out with a StepOne<sup>TM</sup> Real-Time PCR System machine provided 201 with the StepOne software v. 2.3 (Life Technologies, Carlsbad, CA). DNA samples and 202 203 standards were prepared for real-time PCR using the 2× Takara Premix Ex Taq<sup>TM</sup> Perfect 204 Real-Time (Takara Bio Europe, SAS, France), the TaqMan probe (200 nM) and primers (800 nM each) described in Parladé et al. (2013), 5 μL of the template DNA and HPLC water to a 205 206 final reaction volume of 20 µL. Thermocycling profile was 95 °C for 30 s, followed by 40 cycles of 95 °C for 5 s and 60 °C for 34 s. The standard curve was generated from unripe T. 207 melanosporum sporocarps as described in Parladé et al. (2013). 208 209 210 Root colonization by arbuscular mycorrhizal fungi 211 The extent of mycorrhizal colonization in the root systems of the MAPs was assessed under a 212 dissecting microscope using the visual estimate of the percentage of cortex infected by the fungus as described by Giovannetti and Mosse (1980). Root samples were previously clarified 213 214 and stained following the procedure described by Koske and Gemma (1989). 215 Essential oil yield and chemical characterization 216 In August 2022, one sample of plant shoots was taken for each of the following: lavenders 217

growing within brûlés, lavenders outside brûlés, rosemaries within brûlés and rosemaries

outside brûles. Each sample was composed of the shoots of five plants pooled together. The plants were randomly selected. Laboratory-scale hydro-distillation (three replicates of 100 g fresh plant material) was carried out in a Clevenger type apparatus for quality control, according to the European Pharmacopoeia (Navarro-Rocha et al. 2020). The essential oil yield was determined as the volume of oil (mL) extracted from a weight of distilled dry plant (kg). An aliquot of the oil sample was subjected to gas chromatography-mass spectrometry (GC-MS) to determine the essential oil composition. Essential oil constituents are key markers of the oil quality (Lafhal et al. 2016). A 6890 series chromatograph coupled with a 5973N serie mass selective detector (Agilent Technologies, California, USA) was used. The instrument was equipped with a capillary column HP-5MS (Agilent Technologies, California, USA) of 30 m, 0.25 mm i.d., 0.25 µm film thickness and a flow of 1 mL/min with helium as a carrier gas. The oven temperature was 60 °C held for 1 min, then raised at 3 °C/min to 246 °C, for 1 min. The MS used the electron impact mode with an ionization potential of 70 eV and an ion source temperature of 230 °C. The interface temperature was 240 °C. The MS scanning was recorded in full scan mode (40–250 m/z). A ChemStation software was used for controlling the GC-MS system. The amounts of individual compounds were calculated in mode total ion chromatogram (TIC) as the percentage of area of the total GC peak area.

236

237

238

239

240

241

242

243

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

## Data analysis

To assess the effect of intercropping on the MAPs, we analysed the survival of MAPs, the dry weight of their shoot, their shoot height, the percent root colonization by AM fungi and the abundance of *T. melanosporum* extraradical mycelium in the MAP rhizosphere. For each MAP species, the differences between the plants growing within and outside the *brûlé* were tested with a nested model, in which the MAP species was included as a main effects predictor and the *brûlé* / not *brulé* variable was nested within the MAP species. The shoot dry

weight, shoot height and truffle mycelium abundance were tested with general linear models. None of them met all model assumptions (homogeneity of variance, normality and linearity). The shoot dry weight was log-transformed, whereas in the case of shoot height and truffle mycelium, a generalised (gamma) linear model was used instead. The MAP survival and the proportion of MAP roots colonized by AM fungi were analysed with generalised (binomial) linear models. In generalised linear models the fit of the error structure was assessed through overdispersion. Least-squares means tests were used for post-hoc comparisons, with a P=0.05 threshold for statistical significance. The volatile compound profile of the essential oils was analysed with principal components analysis (PCA).

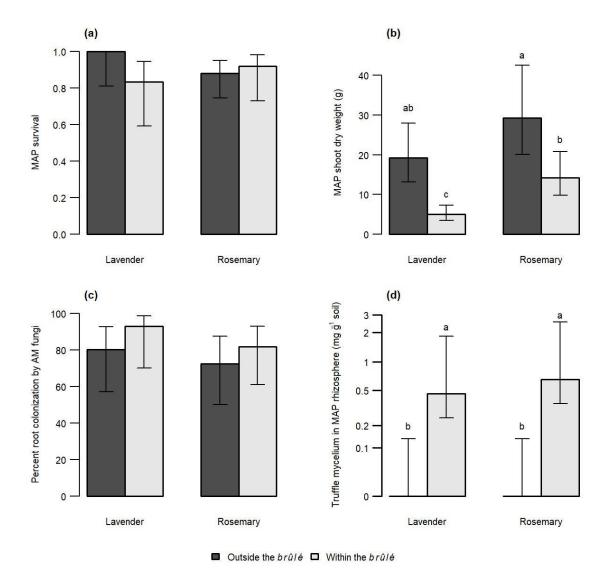
To assess the effect of intercropping on truffle, the abundance of T. melanosporum extraradical mycelium in the bulk soil of the br $\hat{u}$ l $\hat{e}$  was compared to the rhizosphere of MAPs planted in the br $\hat{u}$ l $\hat{e}$ , for each of the two MAPs studied. Data were analysed with a general linear model in which soil position (bulk soil / MAP rhizosphere) was nested within the MAP species. The response variable was log-transformed to meet model assumptions. All analyses were conducted with R and the emmeans package (Lenth 2021; R Core Team 2022).

# Results

261 Effects on aromatic plants

Six months after planting, neither the survival of lavender nor that of rosemary was significantly affected by the  $br\hat{u}l\acute{e}$  (z=-0.01, P-value = 0.99 and z=0.50, P-value = 0.62, respectively, n=108, Fig. 2a). The shoot dry weight was significantly higher outside than inside the  $br\hat{u}l\acute{e}$  for both lavender and rosemary (t=-5.19, P-value < 0.001 and t=-2.78, P-value < 0.010, respectively, n=32, Fig. 2b). The height of the MAP shoots followed a similar pattern, although the decrease of height in  $br\hat{u}l\acute{e}s$  was only significant for lavender and not for

rosemary (Table S1, Fig. S1). The height and the dry weight of MAP shoots showed a significant positive correlation (Spearman's r = 0.74, P-value < 0.001, n = 32).



**Figure 2**. Effect of the truffle  $br\hat{u}l\acute{e}$  on the lavender and rosemary survival (a), shoot dry weight (b), percent root colonization by arbuscular mycorrhizal (AM) fungi (c), and abundance of T. melanosporum mycelium in the aromatic plant (MAP) rhizosphere (d), six months after planting the MAPs (predicted values and 95% confidence interval). Letters indicate significant differences among treatments according to least-squares means tests ( $\alpha$  =

0.05, n = 108 for the survival, n = 32 for shoot dry weight and truffle mycelium, and n = 12 for AM fungi colonization). The y-axis in (d) is represented in log scale.

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

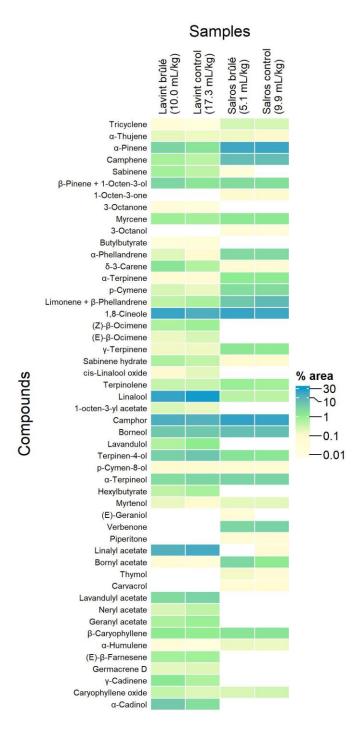
301

277

278

The percent root colonization of the MAPs by AM fungi was not significantly affected by the  $br\hat{u}l\acute{e}$  either in lavender or rosemary (t = 1.1, P-value = 0.29 and t = 0.7, P-value = 0.48, respectively, n = 12, Fig. 2c). On the other hand, the abundance of *T. melanosporum* extraradical mycelium was significantly higher in the rhizosphere of the MAPs inside the  $br\hat{u}l\acute{e}$  than in those outside, for both lavender and rosemary (t = -2.4, P-value = 0.022 and t = -2.5, P-value = 0.019, respectively, n = 32, Fig. 2d). The dry weight of MAP shoots and the abundance of *T. melanosporum* extraradical mycelium in the rhizosphere of the MAPs showed a significant negative correlation (Spearman's r = -0.53, P-value = 0.001, n = 32). However, when the mycelium abundance in the rhizosphere of dead MAPs was compared with that of living MAPs in the same brûlé, no significant differences were found either for lavender or for rosemary (Table S2, Fig. S2). Finally, no significant correlation was found between AM fungi colonization of MAPs and T. melanosporum mycelium abundance in the rhizosphere of these MAPs (Spearman's r = 0.40, P-value = 0.20, n = 12). The extraction of essential oils from the shoots of lavender yielded 10.0 mL kg<sup>-1</sup> dry matter for the plants growing within the *brûlé* and 17.3 mL kg<sup>-1</sup> for those growing outside it. For rosemary, shoots yielded 5.1 mL kg<sup>-1</sup> for the plants growing within the *brûlé* and 9.9 mL kg<sup>-1</sup> for those growing outside. The GC-MS allowed to identify a total of 43 volatile compounds in the lavender oil samples and 34 in the rosemary oil samples (Fig. 3). The most abundant compounds in lavender oil were linalool, 1,8-cineole, linalyl acetate and camphor. For rosemary the most abundant compounds were camphor, 1.8-cineole and  $\alpha$ -pinene, and to a lesser degree borneol, camphene and limonene + β-phellandrene (Fig. 3, Table S3). The PCA for the scaled values of the percentage area explained 83.7% of the total variability with the

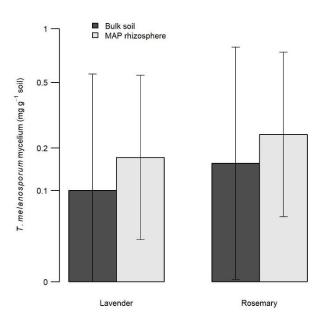
first PCA component, which clearly separated the oil of lavender from that of rosemary (Fig. S3). An additional 11.7% of the variability was explained with the second component. The second component separated the lavender samples growing within and outside the  $br\hat{u}l\acute{e}$  based on 1,8-cineole and the less abundant  $\beta$ -pinene + 1-Octen-3-ol, which presented the more positive loadings with the second PCA component, in relation with their higher abundance within the  $br\hat{u}l\acute{e}$  (Fig. S3). However, the second PCA component could not separate the rosemary samples (Fig. S3).



**Figure 3**. Heatmap for the relative frequency (percentage of area values, indicated by color) of the volatile compounds detected by GC-MS in the essential oil extracted from lavender (Lavint) and rosemary (Salros) cultivated within  $br\hat{u}l\acute{e}s$  and outside them (control) (n = 4). The yield of the essential oil extraction is added next to each sample.

Effect on truffle extraradical mycelium

The effect of intercropping on truffle was assessed by comparing extraradical mycelium abundance in the bulk soil and in the MAP rhizosphere. Within the  $br\hat{u}l\acute{e}$ , the abundance of T. melanosporum extraradical mycelium was not significantly different in the bulk soil and in the rhizosphere of the MAPs, either for lavender or rosemary (t = 0.48, P-value = 0.64 and t = 0.42, P-value = 0.68, respectively, t = 0.48, Fig. 4). Outside the  $br\hat{u}l\acute{e}$ , no truffle extraradical mycelium was found in the bulk soil, whereas in the rhizosphere of MAPs it only appeared in one (6%) of the samples, with less than 0.001 mg g<sup>-1</sup> soil.



**Figure 4**. Effect of intercropping with lavender and rosemary on abundance of T. melanosporum extraradical mycelium: comparison between the bulk soil of the  $br\hat{u}l\acute{e}$  and the rhizosphere of the aromatic plants (MAPs) planted in the  $br\hat{u}l\acute{e}$ , six months after planting (predicted values of the linear model and 95% confidence interval,  $\alpha = 0.05$ , n = 24). The y-axis is represented in log scale.

# **Discussion**

Intercropping with MAPs has the potential to promote ecosystem services in Mediterranean orchards if crop complementarity or facilitation is possible (Durán-Zuazo et al. 2008; Song et al. 2010). The sustained coexistence of the truffle – MAPs intercropping is potentially challenged in adult stages of the truffle plantation due to interference between crops stemming from *brûlé* spread (Martin-Chave 2019). Developing integrative practices that enhance crop complementarity beyond the early stage of the truffle plantation requires scientific information on the ecological relationships between truffle and MAPs (Geoffroy et al. 2018). In this study, we hypothesised that the truffle – MAPs intercropping would be associated with reduced survival and growth of MAPs in the brûlé, higher yield of essential oils of MAPs and altered volatile compounds profiles. We also hypothesised that the competition between T. melanosporum and VA fungi would result in lower colonization of MAP roots by AM fungi and lower abundance of truffle mycelium in the *brûlé*. The study showed a strong negative relation of brûlé soil with the shoot growth of lavender and rosemary during their first vegetative period in the field, although not with their survival. It also suggested a negative relation with the yield of essential oils. This could affect the productivity of lavender and rosemary because their shoots begin to be harvested from the second year (More et al. 2010). It would be interesting to assess how this growth decrease could have a negative influence on other ecosystem services of MAPs such as carbon sequestration or enhanced pollinator populations. The *brûlé* of *T. melanosporum* heavily inhibits the germination and growth of many herbaceous and shrub species (González-Armada et al. 2010; Pacioni 1991; Taschen et al. 2020). This has generally been attributed to the allelopathic effect of truffle metabolites and the endophytic interaction of truffle mycelium with the roots of these plants (Schneider-Maunoury et al. 2020; Streiblová et al. 2012). In our study, T. melanosporum mycelium was practically absent outside the brûlé, and the shoot growth of MAPs was negatively correlated

333

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

to the abundance of *T. melanosporum* mycelium in their rhizosphere. However, we cannot reject the hypothesis that MAP growth decrease is partly caused by root competition between MAPs and oaks, or by the competition between the fungi associated to their respective roots. Previous studies have frequently found a negative relationship between ECM and AM fungi (Chen et al. 2000; Knoblochová et al. 2017). However, we found no negative relationship of AM fungi colonization with *brûlé* occurrence or abundance of *T. melanosporum* mycelium, thus suggesting that MAP growth decrease is not related to deficient AM colonization. Despite previous studies suggesting reduced survival of herbaceous and shrub species in T. melanosporum brûlés (González-Armada et al. 2010; Pacioni 1991), we did not find such an effect in the planted MAPs during their first vegetative period in the field. We hypothesise that this is due to the MAPs being introduced as plantlets, whereas weeds usually arrive at brûlés as seeds. Taschen et al. (2020) showed that truffle mycelium inhibited germination of weed seeds in rhizotron trials. The recently germinated seedlings are likely much more sensitive to the *brûlé* effect than the bareroot lavender or the containerised rosemary seedlings. This is supported by the lack of correlation between MAP mortality and abundance of T. melanosporum mycelium, which suggests that mortality was more related to the preplanting vigour or physiological conditions of the seedlings. With regard to the essential oils of the MAPs, we unexpectedly found lower yields in plants growing within brûlés, where the conditions for MAPs are apparently more stressful. In water stress experiments, it is frequent that Lavandula and Salvia species increase essential oils yield as a mean to protect themselves from water stress (Chrysargyris et al. 2016; Sarmoum et al. 2019). This would suggest more intense stress conditions in the brûlé or different biochemical/physiological mechanisms from those triggered by water stress (Kleinwächter and Selmar 2015). Previous studies also found that water stress altered the composition of essential oil, particularly in α-pinene, limonene, 1,8-cineole and camphor (Chrysargyris et al.

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

2016; Kulak 2020). Our study found an increased abundance of 1,8-cineole in lavenders growing within the brûlé, in agreement with Chrysargyris et al. (2016) and Kulak (2020) who found that water stress increased the abundance of this compound. Besides, the PCA shows that the *brûlé* involved a more substantial modification of the volatile compounds profile for lavender than for rosemary. This suggests a higher potential for influencing the essential oil commercial quality, although this should be confirmed in subsequent years (Kivrak 2018). Our results did not show that the mycelium of T. melanosporum was negatively affected by MAP presence in the *brûlé*, the soil where most truffle fruitbodies will be produced. The finding that truffle mycelium abundance was similar in the MAP rhizosphere and the bulk soil was unexpected, because MAP roots are colonised by AM fungi and previous studies have shown negative relationships between AM and ECM fungi (Chen et al. 2000; Knoblochová et al. 2017). This is all the more remarkable considering that MAPs show high levels of essential oils with biocidal properties (Garzoli et al. 2021; Valcárcel et al. 2021). Interestingly, Taschen et al. (2020) found that some non-ECM plants promoted truffle mycelium growth in rhizotron experiments, including Thymus vulgaris L., a MAP species. They hypothesised that this could be related to enhanced nutrition (through parasitism of non-ECM plants or thanks to rootassociated microorganisms increasing soil nutrients availability). Another possible explanation would be a change in the distribution pattern of the oak fine roots in response to competition with non-ECM plants. Previous studies showed that oak fine roots tended to concentrate in soil layers where weed roots grew (Cubera et al. 2012; Gómez-Molina et al. 2020). Finally, we cannot rule out that truffle mycelium growth was boosted by the change in soil conditions: soil was locally disturbed and loosened to plant MAPs and then irrigated for a while. It would be interesting to monitor whether further growth of MAPs in subsequent years will affect T. melanosporum mycelium. Previous studies show that T. melanosporum

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

mycelium is more abundant in productive than in non-productive trees, thus suggesting it influences truffle orchard productivity (Parladé et al. 2013; Queralt et al. 2017). In conclusion, we found that the shoot growth and essential oil yield of lavender and rosemary seedlings was impaired during their first vegetative period growing in truffle brûlé soils, which are characterized by the presence of T. melanosporum as extraradical mycelium. On the other hand, the abundance of T. melanosporum extraradical mycelium was not affected by the occurrence of lavender and rosemary seedlings in the brûlé. We showed that the occurrence of T. melanosporum mycelium outside the brûlé was very limited, and the normal growth of the MAPs was not apparently disturbed, thus supporting the potential of truffle – MAPs intercropping during the early stage of the truffle plantation. Truffle orchards are typically planted at broad frameworks (e. g.  $6 \times 6$  m), thus allowing three rows of lavender or rosemary to be installed between tree rows, separated at least two meters from trees (Martin-Chave 2019). During the first lavender or rosemary cultivation cycle (8-10 years), the contact between tree and MAP roots is expected to be very limited. The challenges will likely begin in a second MAP cultivation cycle due to the *brûlé* spread, with a MAP growth reduction that will reduce its essential oil production and may also affect oil quality and MAP ecosystem services such as carbon sequestration or enhancement of pollinator populations. It would be interesting to monitor and quantify the findings of this work throughout the entire cultivation cycle of lavender and rosemary, to exactly evaluate the economic viability of intercropping. In this monitoring, productive aspects should be addressed, such as the changes in the yield and quality of the MAP essential oils or the increase in truffle fruitbody yield and quality suggested by some MAP growers (Martin-Chave 2019). The latter could be related to changes in soil properties, and particularly in soil microbial populations (Geoffroy et al. 2018). It would also be interesting to investigate a potential role of MAPs (and their essential oils) in

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

431	the control of the pest insect Leiodes cinnamomeus Panzer, which is the main problem
432	nowadays in many truffle plantations in eastern Spain (Julià et al. 2023).
433	
434	Declaration of Competing Interest
435	The authors declare that they have no known competing financial interests or personal
436	relationships that could have appeared to influence the work reported in this paper.
437	
438	CRediT authorship contribution statement
439	Conceptualization and methodology, S.GB., S.S., E.GM., J.P. and J.NR.; investigation,
440	E.GM., V.B., J.P., M.A.S., J.NR. and S.S.; formal analysis, S.GB.; writing – original
441	draft, S.GB., S.S., J.P. and J.NR.; writing – review and editing, all authors.
442	
443	Funding
444	This research was supported by the Collaboration Agreement for the Operation of CIET
445	(funded by Diputación Provincial de Huesca, with the participation of CITA, Comarca de
446	Ribagorza and Ayuntamiento de Graus), the Spanish Ministry of Science, Innovation and
447	Universities [grant number RTI2018-093907-B-C21/C22], and European Union
448	NextGenerationEU funding [grant number PRTR-C17.I1].
449	
450	Data availability
451	The datasets generated during and/or analysed during the current study are available from the
452	corresponding author on reasonable request.
453	
454	References

Andrés-Alpuente A, Sánchez S, Martín M, et al (2014) Comparative analysis of different

456	methods for evaluating quality of Quercus ilex seedlings inoculated with Tuber
457	melanosporum. Mycorrhiza 24:S29–S37. https://doi.org/10.1007/s00572-014-0563-x
458	Aumeeruddy-Thomas Y, Taschen E, Richard F (2016) Taming the black tuffle (Tuber
459	melanosporum). Safeguarding Mediterranean food and ecological webs. Mediterr Reg
460	Under Clim Chang A Sci Updat 533–542
461	Benucci GMN, Bonito GM (2016) The truffle microbiome: species and geography effects on
462	bacteria associated with fruiting bodies of hypogeous Pezizales. Microb Ecol 72:4-8.
463	https://doi.org/10.1007/s00248-016-0755-3
464	Brunori E, Maesano M, Moresi FV, et al (2020) The hidden land conservation benefits of
465	olive-based (Olea europaea L.) landscapes: An agroforestry investigation in the southern
466	Mediterranean (Calabria region, Italy). L Degrad Dev 31:801-815.
467	https://doi.org/10.1002/ldr.3484
468	Chen X, Song B, Yao Y, et al (2014) Aromatic plants play an important role in promoting soil
469	biological activity related to nitrogen cycling in an orchard ecosystem. Sci Total Environ
470	472:939–946. https://doi.org/10.1016/j.scitotenv.2013.11.117
471	Chen YL, Brundrett MC, Dell B (2000) Effects of ectomycorrhizas and vesicular-arbuscular
472	mycorrhizas, alone or in competition, on root colonization and growth of Eucalyptus
473	globulus and E. urophylla. New Phytol 146:545–555. https://doi.org/10.1046/j.1469-
474	8137.2000.00663.x
475	Chrysargyris A, Laoutari S, Litskas VD, et al (2016) Effects of water stress on lavender and
476	sage biomass production, essential oil composition and biocidal properties against
477	Tetranychus urticae (Koch). Sci Hortic (Amsterdam) 213:96–103.
478	https://doi.org/10.1016/j.scienta.2016.10.024
479	Cubera E, Moreno G, Solla A, Madeira M (2012) Root system of <i>Quercus suber</i> L. seedlings
480	in response to herbaceous competition and different watering and fertilisation regimes.

481	Agrofor Syst 85:205–214. https://doi.org/10.1007/s10457-012-9492-x
482	De Franchis L (2003) Threats to soils in Mediterranean countries. UNEP, Valbonne (France)
483	Demestihas C, Plénet D, Génard M, et al (2017) Ecosystem services in orchards. A review.
484	Agron Sustain Dev 37:. https://doi.org/10.1007/s13593-017-0422-1
485	Demestihas C, Plénet D, Génard M, et al (2019) A simulation study of synergies and tradeoffs
486	between multiple ecosystem services in apple orchards. J Environ Manage 236:1–16.
487	https://doi.org/10.1016/j.jenvman.2019.01.073
488	Durán-Zuazo VH, Rodríguez-Pleguezuelo CR, Francia-Martínez JR, et al (2008) Benefits of
489	plant strips for sustainable mountain agriculture. Agron Sustain Dev 28:497-505.
490	https://doi.org/10.1051/agro:2008020
491	Fanlo M, Melero R, Moré E, Cristóbal R (2009) Cultivo de plantas aromáticas, medicinales y
492	condimentarias en Cataluña. Centre Tecnològic Forestal de Catalunya, Solsona (Spain)
493	Garcia-Barreda S, Sánchez S, Marco P, Serrano-Notivoli R (2019) Agro-climatic zoning of
494	Spanish forests naturally producing black truffle. Agric For Meteorol 269–270:231–238.
495	https://doi.org/10.1016/j.agrformet.2019.02.020
496	Garzoli S, Laghezza-Masci V, Franceschi S, et al (2021) Headspace/GC-MS analysis and
497	investigation of antibacterial, antioxidant and cytotoxic activity of essential oils and
498	hydrolates from Rosmarinus officinalis L. and Lavandula angustifolia Miller. Foods
499	10:1768
500	Geoffroy A, Richard F, Sanguin H (2018) Impact of intercropping cultures on truffle
501	production and soil microbial communities in Mediterranean oak orchards. In:
502	International conference on ecological sciences. Rennes (France)
503	Giovannetti M, Mosse B (1980) An evaluation of techniques for measuring vesicular
504	arbuscular mycorrhizal infection in roots. New Phytol 84:489-500.
505	https://doi.org/10.1111/J.1469-8137.1980.TB04556.X

506	Gómez-Molina E, Sánchez S, Parladé J, et al (2020) Glyphosate treatments for weed control
507	affect early stages of root colonization by Tuber melanosporum but not secondary
508	colonization. Mycorrhiza 30:725–733. https://doi.org/10.1007/S00572-020-00990-8
509	González-Armada B, De Miguel AM, Cavero RY (2010) Ectomycorrhizae and vascular
510	plants growing in brûlés as indicators of below and above ground microecology of black
511	truffle production areas in Navarra (Northern Spain). Biodivers Conserv 19:3861–3891.
512	https://doi.org/10.1007/s10531-010-9935-5
513	Julià I, Morton A, Garcia-del-Pino F (2023) The development of the truffle beetle Leiodes
514	cinnamomeus at low temperature, a determining factor for the susceptibility of adults and
515	larvae to entomopathogenic nematodes. Biol Control 180:105197.
516	https://doi.org/10.1016/J.BIOCONTROL.2023.105197
517	Kivrak S (2018) Essential oil composition and antioxidant activities of eight cultivars of
518	lavender and lavandin from western Anatolia. Ind Crops Prod 117:88–96.
519	https://doi.org/10.1016/J.INDCROP.2018.02.089
520	Kleinwächter M, Selmar D (2015) New insights explain that drought stress enhances the
521	quality of spice and medicinal plants: potential applications. Agron Sustain Dev 35:121-
522	131. https://doi.org/10.1007/S13593-014-0260-3/TABLES/1
523	Knoblochová T, Kohout P, Püschel D, et al (2017) Asymmetric response of root-associated
524	fungal communities of an arbuscular mycorrhizal grass and an ectomycorrhizal tree to
525	their coexistence in primary succession. Mycorrhiza 27:775–789.
526	https://doi.org/10.1007/s00572-017-0792-x
527	Koske RE, Gemma JN (1989) A modified procedure for staining roots to detect VA
528	mycorrhizas. Mycol Res 92:486–488. https://doi.org/10.1016/S0953-7562(89)80195-9
529	Kulak M (2020) Recurrent drought stress effects on essential oil profile of Lamiaceae plants:
530	An approach regarding stress memory. Ind Crops Prod 154:112695.

531	https://doi.org/10.1016/J.INDCROP.2020.112695
532	Lafhal S, Vanloot P, Bombarda I, et al (2016) Identification of metabolomic markers of
533	lavender and lavandin essential oils using mid-infrared spectroscopy. Vib Spectrosc
534	85:79–90. https://doi.org/10.1016/J.VIBSPEC.2016.04.004
535	Lagacherie P, Alvaro-Fuentes J, Annabi M, et al (2018) Managing Mediterranean soil
536	resources under global change: expected trends and mitigation strategies. Reg Environ
537	Chang 18:663–675. https://doi.org/10.1007/s10113-017-1239-9
538	Le Tacon F, Zeller B, Plain C, et al (2013) Carbon transfer from the host to Tuber
539	melanosporum mycorrhizas and ascocarps followed using a 13C pulse-labeling
540	technique. PLoS One 8:e64626. https://doi.org/10.1371/journal.pone.0064626
541	Lenth R V (2021) emmeans: Estimated Marginal Means, aka Least-Squares Means
542	Martin-Chave A (2019) Produire de la truffe en agroforesterie: pratiques traditionnelles te
543	expérimentations dans le Sud-Est. Scoop-Agroof, Anduze (France)
544	Mello A, Balestrini R (2018) Recent insights on biological and ecological aspects of
545	ectomycorrhizal fungi and their interactions. Front Microbiol 9:1-13.
546	https://doi.org/10.3389/fmicb.2018.00216
547	More E, Fanlo M, Melero R, Cristobal R (2010) Guía para la producción sostenible de plantas
548	aromáticas y medicinales
549	Morugán-Coronado A, Linares C, Gómez-López MD, et al (2020) The impact of
550	intercropping, tillage and fertilizer type on soil and crop yield in fruit orchards under
551	Mediterranean conditions: A meta-analysis of field studies. Agric Syst 178:102736.
552	https://doi.org/10.1016/j.agsy.2019.102736
553	Navarro-Rocha J, Andrés MF, Díaz CE, et al (2020) Composition and biocidal properties of
554	essential oil from pre-domesticated Spanish Satureja montana. Ind Crops Prod
555	145:111958. https://doi.org/10.1016/J.INDCROP.2019.111958

556	Pacioni G (1991) Effects of <i>Tuber</i> metabolites on the rhizospheric environment. Mycol Res
557	95:1355–1358. https://doi.org/10.1016/S0953-7562(09)80384-5
558	Parladé J, De la Varga H, De Miguel AM, et al (2013) Quantification of extraradical
559	mycelium of Tuber melanosporum in soils from truffle orchards in northern Spain.
560	Mycorrhiza 23:99-106. https://doi.org/10.1007/S00572-012-0454-Y
561	Plattner I, Hall I (1995) Parasitism of non-host plants by the mycorrhizal fungus <i>Tuber</i>
562	melanosporum. Mycol Res 99:1367–1370
563	Queralt M, Parladé J, Pera J, De Miguel AM (2017) Seasonal dynamics of extraradical
564	mycelium and mycorrhizas in a black truffle (Tuber melanosporum) plantation.
565	Mycorrhiza 27:565–576
566	R Core Team (2022) R: a language and environment for statistical computing
567	Sarmoum R, Haid S, Biche M, et al (2019) Effect of salinity and water stress on the essential
568	oil components of rosemary (Rosmarinus officinalis L.). Agronomy 9:214.
569	https://doi.org/10.3390/AGRONOMY9050214
570	Schneider-Maunoury L, Deveau A, Moreno M, et al (2020) Two ectomycorrhizal truffles,
571	Tuber melanosporum and T. aestivum, endophytically colonise roots of non-
572	ectomycorrhizal plants in natural environments. New Phytol 225:2542-2556.
573	https://doi.org/10.1111/nph.16321
574	Schneider-Maunoury L, Leclercq S, Clément C, et al (2018) Is Tuber melanosporum
575	colonizing the roots of herbaceous, non-ectomycorrhizal plants? Fungal Ecol 31:59-68.
576	https://doi.org/10.1016/j.funeco.2017.10.004
577	Song BZ, Wu HY, Kong Y, et al (2010) Effects of intercropping with aromatic plants on the
578	diversity and structure of an arthropod community in a pear orchard. BioControl 55:741-
579	751. https://doi.org/10.1007/s10526-010-9301-2
580	Splivallo R, Ottonello S, Mello A, Karlovsky P (2011) Truffle volatiles: from chemical

581	ecology to aroma biosynthesis. New Phytol 189:688–699. https://doi.org/10.1111/j.1469-
582	8137.2010.03523.x
583	Streiblová E, Gryndlerová H, Gryndler M (2012) Truffle brûlé: An efficient fungal life
584	strategy. FEMS Microbiol Ecol 80:1-8. https://doi.org/10.1111/j.1574-
585	6941.2011.01283.x
586	Taschen E, Sauve M, Vincent B, et al (2020) Insight into the truffle brûlé: tripartite
587	interactions between the black truffle (Tuber melanosporum), holm oak (Quercus ilex)
588	and arbuscular mycorrhizal plants. Plant Soil 446:577-594.
589	https://doi.org/10.1007/s11104-019-04340-2
590	Valcárcel F, Olmeda AS, González MG, et al (2021) Acaricidal and insect antifeedant effects
591	of essential oils from selected aromatic plants and their main components. Front Agron
592	3:1–12. https://doi.org/10.3389/fagro.2021.662802
593	Zhang Y, Han M, Song M, et al (2021) Intercropping with aromatic plants increased the soil
594	organic matter content and changed the microbial community in a pear orchard. Front
595	Microbiol 12:1–15. https://doi.org/10.3389/fmicb.2021.616932
596	